1 1. A nerve regeneration conduit comprising a porous biocompatible support 2 comprising an inner surface and an outer surface, the support being in the form of a roll 3 such that a cross section of the roll approximates a spiral spanning from 8 to 40 rotations, 4 with the outer surface of the support facing outward, relative to the origin of the spiral. 1 2. The nerve regeneration conduit of claim 1, wherein the support has a thickness 2 of 5 to 200 µm. 1 3. The nerve regeneration conduit of claim 1, wherein the support has a thickness 2 of 10 to 100 μm. 1 4. The nerve regeneration conduit of claim 1, wherein the support comprises a 2 biological material. 1 5. The nerve regeneration conduit of claim 4, wherein the biological material is 2 small intestinal submucosa. 1 6. The nerve regeneration conduit of claim 1, wherein the support comprises a 2 synthetic polymer. 1 7. The nerve regeneration conduit of claim 1, wherein the support is 2 bioresorbable. 1 8. The nerve regeneration conduit of claim 6, wherein the synthetic polymer is 2 selected from the group consisting of polyhydroxyalkanoates, e.g., polyhydroxybutyric 3 acid; polyesters, e.g., polyglycolic acid (PGA); copolymers of glycolic acid and lactic 4 acid (PLGA); copolymers of lactic acid and ε-aminocaproic acid; polycaprolactones; 5 polydesoxazon (PDS); copolymers of hydroxybutyric acid and hydroxyvaleric acid; 6 polyesters of succinic acid; polylactic acid (PLA); cross-linked hyaluronic acid; 7 poly(organo)phosphazenes; biodegradable polyurethanes; and PGA cross-linked to 8 collagen.

1 9. The nerve regeneration conduit of claim 1, further comprising a layer of cells 2 adhered to the inner surface of the support. 1 10. The nerve regeneration conduit of claim 9, wherein the cells are Schwann 2 cells or olfactory ensheathing glial cells. 1 11. The nerve regeneration conduit of claim 10, wherein the layer contains from 2 15,000 to 165,000 Schwann cells per millimeter of conduit length. 1 12. The nerve regeneration conduit of claim 11, wherein the layer contains from 2 20,000 to 40,000 Schwann cells per millimeter of conduit length. 1 13. The nerve regeneration conduit of claim 9, further comprising a layer of 2 extracellular matrix material on the support. 1 14. The nerve regeneration conduit of claim 1, further comprising a hydrogel 2 layer. 1 15. The nerve regeneration conduit of claim 14, wherein the hydrogel layer has a 2 thickness of 5 to 120 µm. 1 16. The nerve regeneration conduit of claim 15, wherein the hydrogel layer has a 2 thickness of 10 to 50 µm. 1 17. The nerve regeneration conduit of claim 14, wherein the hydrogel layer 2 comprises a polymer selected from the group consisting of fibrin glues, Pluronics<sup>®</sup>. 3 polyethylene glycol (PEG) hydrogels, agarose gels, PolyHEMA (poly 2-4 hydroxyethylmethacrylate) hydrogels, PHPMA (poly N-(2-hydroxypropyl) methacrylamide) hydrogels, collagen gels, Matrigel<sup>®</sup>, chitosan gels, gel mixtures (e.g., of 5 6 collagen, laminin, fibronectin), alginate gels, and collagen-glycosaminoglycan gels.

1 18. The nerve regeneration conduit of claim 1, further comprising a multiplicity 2 of microspheres. 1 19. The nerve regeneration conduit of claim 18, wherein the microspheres are 2 immobilized in a hydrogel layer. 1 20. The nerve regeneration conduit of claim 14, wherein the hydrogel layer 2 comprises a neurotrophic agent. 1 21. The nerve regeneration conduit of claim 18, wherein the microspheres 2 comprise a neurotrophic agent. 1 22. The nerve regeneration conduit of claim 18, wherein the microspheres have a 2 diameter of 1 to 150 µm. 1 23. The nerve regeneration conduit of claim 18, wherein the microspheres 2 comprise a material selected from the group consisting of a polyhydroxyalkanoate, a 3 polyester, a copolymer of glycolic acid and lactic acid (PLGA), a copolymer of lactic 4 acid and ε-aminocaproic acid, a polycaprolactones, polydesoxazon (PDS), a copolymer of 5 hydroxybutyric acid and hydroxyvaleric acid, a polyester of succinic acid; and crosslinked hyaluronic acid. 6 1 24. The nerve regeneration conduit of claim 23, wherein the microspheres 2 comprise PLGA having an average molecular weight of 25 kD to 130 kD. 1 25. The nerve regeneration conduit of claim 24, wherein the lactic acid:glycolic 2 acid ratio is approximately 85:15. 1 26. The nerve regeneration conduit of claim 18, wherein the microspheres are 2 arranged in a pattern to facilitate creation of a neurotrophic agent concentration gradient.

1 27. The nerve regeneration conduit of claim 26, wherein the gradient is radial. 1 28. The nerve regeneration conduit of claim 26, wherein the gradient is axial. 1 29. The nerve regeneration conduit of claim 20 or 21, wherein the neurotrophic 2 agent is selected from the group consisting of FK506, αFGF, βFGF, 4-methylcatechol, 3 NGF, BDNF, CNTF, MNGF, NT-3, GDNF, NT-4/5, CM101, inosine, spermine, 4 spermidine, HSP-27, IGF-I, IGF-II, PDGF, ARIA, LIF, VIP, GGF, IL-1, and MS-430. 1 30. The nerve regeneration conduit of claim 20, wherein the hydrogel layer 2 comprises two or more neurotrophic agents. 1 31. The nerve regeneration conduit of claim 21, wherein the microspheres 2 comprise two or more neurotrophic agents. 1 32. The nerve regeneration conduit of claim 31, wherein the neurotrophic agents 2 are in separate microspheres. 1 33. The nerve regeneration conduit of claim 31, wherein two or more 2 neurotrophic agents are in a single microsphere. 1 34. A method of manufacturing a nerve regeneration conduit, the method 2 comprising providing a porous biocompatible support comprising an inner surface and an 3 outer surface; and forming the support into a roll such that a cross section of the roll 4 approximates a spiral spanning from 8 to 40 rotations, with the outer surface of the 5 support facing outward, relative to the origin of the spiral.

16

support prior to forming the support into the roll.

35. The method of claim 34, further comprising culturing a layer of cells on the

1

2

1 36. The method of claim 34, further comprising depositing a hydrogel layer on 2 the support before forming the support into a roll. 1 37. The method of claim 34, further comprising incorporating a multiplicity of 2 microspheres into the conduit. 1 38. The method of claim 37, wherein the microspheres comprise a neurotrophic 2 agent. 1 39. A method of facilitating regeneration of a transected nerve across a nerve gap 2 defined by a proximal end of the transected nerve and a distal end of the transected nerve. 3 the method comprising coapting the proximal end of the transected nerve to a first end of 4 the conduit of claim 1, and coapting the distal end of the transected nerve to a second end 5 of the conduit. 1 40. A method of facilitating regeneration of a crushed nerve, the method 2 comprising providing a porous biocompatible support comprising an inner surface and an 3 outer surface; culturing a layer of cells on the support; and rolling the support around the 4 crushed nerve. 1 41. The method of claim 40, further comprising depositing a hydrogel layer on 2 the support before rolling the support around the crushed nerve. 1 42. The method of claim 40, further comprising incorporating a multiplicity of 2 neurotrophic agent-laden microspheres into the conduit. 1 43. The nerve regenerating conduit of claim 14, wherein the hydrogel further 2 comprises cells. 1 44. The nerve regenerating conduit of claim 1, wherein the support further 2 comprises spacer members extending from the inner surface of the support.

- 1 45. The nerve regenerating conduit of claim 1, wherein the support is loaded with 2 one or more neurotrophins.
- 1 46. The nerve regenerating conduit of claim 45, wherein the one or more
- 2 neurotrophins are distributed in a gradient in the support.